

**Supporting Information**

**Lipophilic Bisphosphonates as Dual Farnesyl/Geranylgeranyl Diphosphate**

**Synthase Inhibitors: An X-ray and NMR investigation**

By

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Table shows a compilation of the outputs of a combinatorial descriptor search that relates cell growth inhibition ( $\text{pIC}_{50}$  (cell)) with GGPPS inhibition plus two other descriptors (B, C).

$$\text{pIC}_{50}(\text{cell}) = a \cdot \text{pIC}_{50}(\text{enzyme}) + b \cdot B + c \cdot C + d$$

where a-d are linear regression coefficient and B, C are chosen from 230 descriptors in MOE and FPPS. The explanation of the meanings of each of the top-10 descriptor sets are given in the MOE manual and ref. 19 in the Text. GGPPS inhibition is the dominant descriptor in most cases, consistent with the high correlation between GGPPS inhibition and cell growth inhibition.

The following gives a summary of the  $R^2$  values based on the combinatorial descriptor search using the top-ranked combination.

Target	Cell line	$R^2$ , Enzyme vs. Cell	$R^2$ , Training Set	$R^2$ , Test Set <sup>a</sup>	Descriptors used
GGPPS	MCF-7	0.60	0.78	0.73	124

<sup>a</sup> Predicted values from leave-two-out cross validated models.



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Target	Cell line	$R^2$ , Enzyme vs. Cell	$R^2$ , Training Set	$R^2$ , Test Set <sup>a</sup>	Descriptors used
GGPPS	NCI-H460	0.54	0.77	0.79	124

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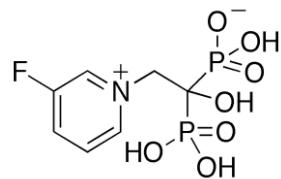
Target	Cell line	$R^2$ , Enzyme vs. Cell	$R^2$ , Training Set	$R^2$ , Test Set <sup>a</sup>	Descriptors used
GGPPS	SF-268	0.56	0.78	0.66	124

<sup>a</sup> Predicted values from leave-two-out cross validated models.

**Table S5:** Data collection and refinement statistics for BPH-461<sup>a</sup> bound to human FPPS.

(PDB: 2OPM)

<u>Crystals</u>	461B <sup>a</sup>
<b>Data collection</b>	
Space group	P4 <sub>1</sub> 2 <sub>1</sub> 2
Unit cell dimension (Å)	
<i>a</i> = <i>b</i> , <i>c</i> (Å)	111.783, 66.525
X-ray source	BNL-X29 <sup>b</sup>
Wavelength (Å)	1.1
Resolution (Å)	30-2.40 (2.49-2.40)
No. of reflection observed	204,362
Unique	16,818 (1,525)
Completeness (%)	98.6 (92.0)
<i>R</i> -merge	0.100 (0.412)
<i>I</i> / <i>σI</i>	42.0
Multiplicity	12.2 (9.2)
<b>Refinement statistics</b>	
Resolution range (Å)	30.0-2.40
<i>R</i> -work/ <i>R</i> -free (%)	23.35/26.93
RMSD	
Bond lengths	0.004
Bond angles	1.532
No. of atoms	
Protein	2,708
Ligand	18
PO <sub>4</sub> <sup>3-</sup>	10
Magnesium ion	3
Solvent (water)	130
<i>B</i> average (Å <sup>2</sup> ) of protein	51.20
<i>B</i> average (Å <sup>2</sup> ) of solvents	56.58
<i>B</i> average (Å <sup>2</sup> ) of ligands(Bisphosphonates,Mg <sup>2+</sup> and PO <sub>4</sub> <sup>3-</sup> )	68.93

<sup>a</sup>: BPH-461 is 3-fluoro-1-(2-hydroxy-2,2-bisphosphonoethyl)-pyridinium<sup>b</sup>: Brookhaven National Laboratory**BPH-461**











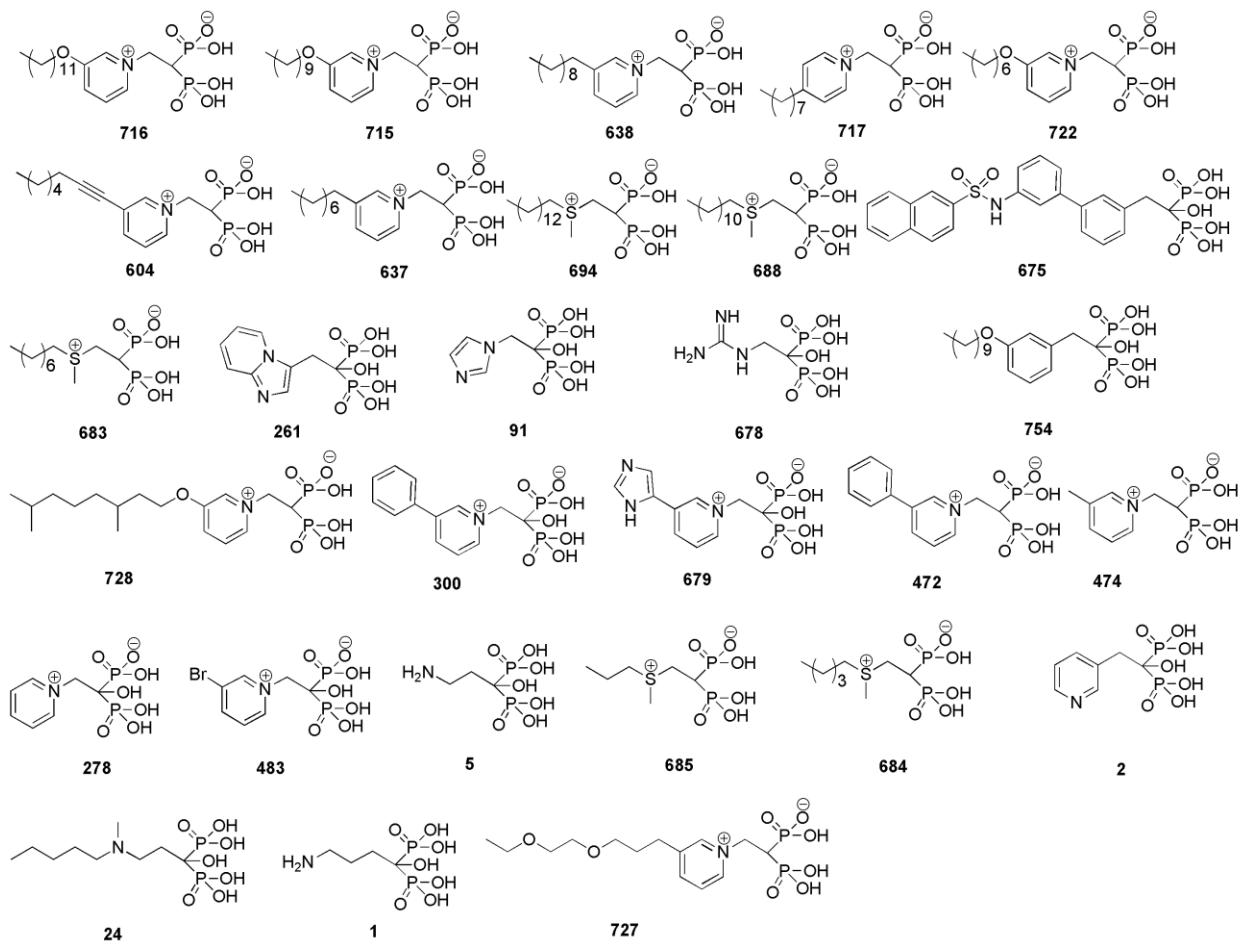
**Table S11:** Isothermal calorimetry results

Cpd ID	$\Delta H$ (kcal/mol)	$\Delta S$ (cal/deg/mol)	$\Delta G$ (kcal/mol)
BPH-527	3.93	42.9	-8.9
BPH-536	4.01	43.1	-8.6
BPH-540	~0		
BPH-541	~0		
BPH-560	3.78	41.5	-8.5
BPH-571	2.49	36.5	-8.3
BPH-678	1.85	35.7	-8.7

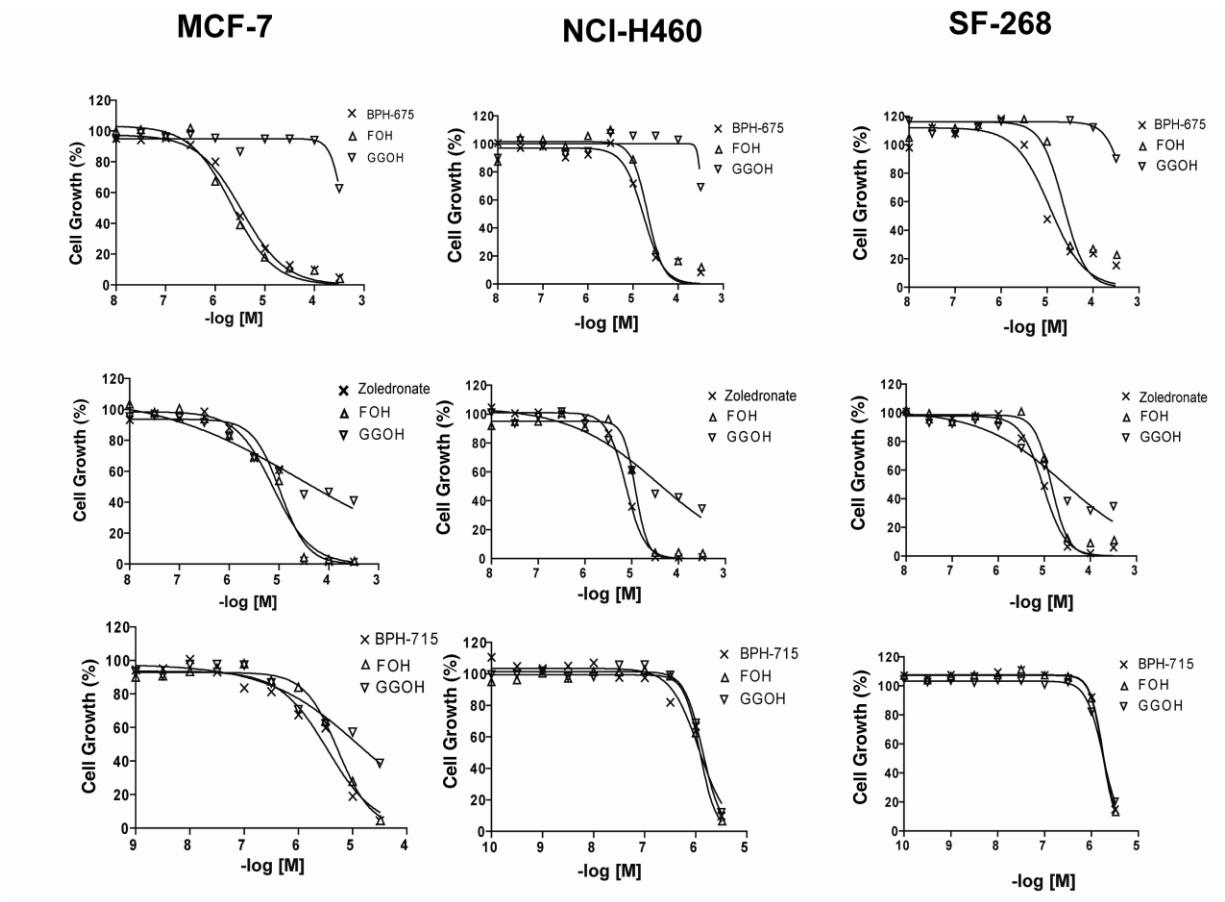




## Supplementary Figures



**Figure S1:** Structures of BPH-series of inhibitors investigated in cell growth inhibition, FPPS and GGPPS enzyme inhibition. Rank ordered in terms of decreasing potency in NCI-H460 cell growth inhibition (see Supporting Table S1).



**Figure S2:** Rescue of cell growth inhibition for all three cell lines; BPH-675, zoledronate and BPH-715 inhibitors; 20  $\mu$ M FOH ( $\Delta$ ) or GGOH ( $\triangledown$ ) concentrations.

## **Supporting Experimental Information**

### NMR sample preparation

*T. brucei* FPPS was expressed in BL21 (DE3) *E. coli* and purified as described previously<sup>1</sup>. Microcrystalline samples for solid state NMR were prepared by serially adding bisphosphonate, IPP and Mg<sup>2+</sup> in a ratio of 1:1:3. Typically, the starting FPPS protein concentration was between 10 to 20 mg/mL, and dilution during ligand addition was kept to a minimum. After incubating for 1-2 hrs, precipitation of crystalline protein<sup>2</sup> was induced by slowly adding first, β-mercaptoethanol (10 mM), then PEG-3350, to a maximum concentration of 20%. The solution was mixed gently and kept overnight until a white precipitate formed. The sample was transferred into a sealed 1 mL Eppendorf tube and then centrifuged at 3220g for 30 min and subsequently transferred into a 3.2 mm NMR rotor by centrifugation (3220g for 30 minutes). In some cases, precipitation was induced by adding Mg<sup>2+</sup> to 2:2:9 (though this was rare). Typically, microcrystals precipitated out at ~ 10% PEG; at 20% or more PEG, a gel formed that was difficult to pellet. It is best to start with high protein concentration of at least 500 μM volume, and to use concentrated stock solutions of bisphosphonates, IPP, Mg<sup>2+</sup>, so that the protein solution is not diluted.

1. Mao, J.; Gao, Y.G.; Odeh, S.; Robinson, H.; Montalvetti, A.; Docampo, R.; Oldfield, E. *Acta Crystallogr D Biol Crystallogr* **2004**, *60*, 1863-6.
2. Martin, R.W.; Zilm, K.M. *J. Mag. Res.* **2003**, *165*, 162-174.

*Complete ref 5 from the article:*

- (5) Gnant, M.; Mlinertsch, B.; Schippinger, W.; Luschin-Ebengreuth, G.; Postlberger, S.; Menzel, C.; Jakesz, R.; Seifert, M.; Hubalek, M.; Bjelic-Radisic, V.; Samonigg, H.; Tausch, C.; Eidtmann, H.; Steger, G.; Kwasny, W.; Dubsky, P.; Fridrik, M.; Fitzal, F.; Stierer, M.; Rucklinger, E.; Greil, R.; Marth, C. *N. Engl. J. Med.* **2009**, *360*, 679-91.